

Effect of *Mikania glomerata* (Asteraceae) leaf extract combined with anti-venom serum on experimental *Crotalus durissus* (Squamata: Viperidae) envenomation in rats

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Abstract: Crotalic envenomation represents the highest number of deaths when compared to other snakebite envenomations of medical interest. Crotalic venom has important characteristics such as neurotoxicity, myotoxicity, nephrotoxicity, and clotting and hemolytic action. We evaluated the clinical and laboratory aspects of *Crotalus durissus terrificus* experimental envenomation in Wistar rats treated with antivenom and the aqueous extract of the plant *Mikania glomerata*. The animals were divided into three groups: *Group C* (control); *Group VS*—venom and antivenom; *Group VSM*—venom, antivenom and aqueous extract of *M. glomerata*. Crotalic poison caused clinical and laboratory alterations in Wistar mice. Significant clinical alterations were: temperature decrease, edema in the venom inoculated member, sedation and a locomotion decrease in groups VS and VSM when compared with group C. A faster recovery from sedation was observed only for animals of group VSM when compared to VS. There was an increase in the number of leukocytes, neutrophils and creatine kinase in the VS and VSM groups, compared to group C. Wistar rats showed a high resistance to crotalic venom. Additional studies with different doses, time of treatment, different administration methods and histopathological and immunological studies are necessary to understand the action of *M. glomerata* in crotalic accidents. Rev. Biol. Trop. 57 (4): 929-937. Epub 2009 December 01.

Keywords: *Crotalus durissus terrificus*, *Mikania glomerata*, antivenom, rat, venom.

Crotalus durissus terrificus snakebites are the second most common cause of casualties in Brazil, and the number one cause of death (Ministério da Saúde 2005, Ministério da Saúde 2006). Crotalic venom has neurotoxic (Vital-Brazil 1972, Amaral *et al.* 1991, Jorge & Ribeiro 1992), systemic myotoxic, and nefrotoxic actions (Amaral *et al.* 1986, Azevedo-Marques *et al.* 1985, Castro 2006). It also changes blood clotting (Amaral *et al.* 1988) and causes hemolysis *in vitro* (Rosenfeld *et al.* 1962, Kelen *et al.* 1962).

Factors like animal species, age, weight, general conditions of the individual organism, amount of injected venom, bite site and time from the envenomation until treatment, contribute to the level of gravity of the envenomation (Murtaugh & Kaplan 1992, Pinho & Pereira 2001).

Serum therapy is the regular treatment, but it is of great importance to find supplementary treatments that might reduce the intensity and gravity of clinical changes, reducing not only the incidence of secondary complications, but

also the number of deaths (Ferreira *et al.* 1992, Pereira *et al.* 1994, Melo *et al.* 1994, Ministério da Saúde 2001, Maiorano *et al.* 2005).

Some plants with antivenom properties are mentioned in the literature (Soares *et al.* 2004, Soares *et al.* 2005), and the *Mikania glomerata* species (F. Asteracea), popularly known as “guaco”, was used in this study. Its extract major component is Cumarine (1,2-benzopirene) that contributes to its antivenom activity (Leite *et al.* 1993). The literature mentions that such plants inhibit the action of several snake venoms. Besides, it is a rich source of potential inhibitors of PLA2, metalloproteases and serine proteases, which are enzymes related to the clinical signs observed in humans and animals (Maiorano *et al.* 2005). This study evaluated the clinical and laboratory aspects of *Crotalus durissus terrificus* experimental envenomation in Wistar rats treated with antivenom and the aqueous extract of *Mikania glomerata*.

MATERIAL AND METHODS

The experimental protocol was approved by the Ethics Committee of University of Western São Paulo, UNOESTE, Presidente Prudente, Brazil (036/06).

Experimental groups: Fifty-four females of the *Rattus norvegicus* species (Berkenhout 1769) of the *Wistar* rat line, weighting between 200 and 300g, were divided into three experimental groups of 18 animals per group. The groups were divided as follows: group VS (venom+serum) received 10mg/kg of crotalic venom via intramuscular injection in the rear left limb and antivenom (Vencofarma®) intraperitoneally at the manufacturer’s recommended dose in order to neutralize the amount of inoculated venom; group VSM (venom+serum+*M. glomerata*) received the same dose of venom and antivenom as Group VS, and an aqueous leaf extract from the plant at a 10% concentration (orally) every hour starting after venom inoculation until a total of five doses was reached. Animals were evaluated at three different times: M1–30 minutes after venom

inoculation; M2–6 hours after venom inoculation; M3–24 hours after venom inoculation.

Animals were housed in stainless steel cages and maintained under standard conditions (12h light/12h dark cycle, constant temperature of 22°C and humidity of 55%). Filtered water and rations were offered *ad libitum*.

Venom: Venom was provided by the Center for the Study of Venoms and Venomous Animals, CEVAP-UNESP, Botucatu, São Paulo, Brazil. Lyophilized venom was diluted in sterile saline solution at a 5mg/mL concentration and inoculated via intramuscular injection in the animal’s rear left limb (lateral face of the thigh).

Antiophidic serum: The bothropic-crotalic serum used was from Vencofarma Laboratory serum (Vencofarma®); each flask of 10mL of serum neutralizes 10mg of crotalic venom. It was administered intraperitoneally six hours after venom inoculation.

Aqueous leaf extract of *M. glomerata*: The extract was prepared at the Pharmacology Laboratory of Adamantinenses Integrated School, FAI, Adamantina, SP, Brazil. The leaves of “guaco” (*M. glomerata*) were collected from plants that were grown in an area that was constantly supervised and checked. The extract preparation followed the Brazilian Pharmacopeia protocol (Silva 1929). The aqueous solution was used at 10% concentration and was intragastrically administered (*gavage*) in 10mg/kg doses, determined in pilot experiments, starting soon after the crotalic venom inoculation and repeated every hour until 5 doses had been given. The plant was identified by Professor Nelson Barbosa Machado Neto of the Universidade do Oeste Paulista-UNOESTE and a voucher specimen (16838) of the plant has been deposited at the Department of Botânica-IBB-UNESP Botucatu, SP.

Physical exam: The physical exam consisted of the evaluation of heart frequency for the inspection of beats per minute (bpm),

breathing frequency for the inspection of the movements per minute (mpm), and temperature was determined with a rectal digital thermometer. The progress of edema was evaluated with a pachymeter after the injection of crotalic venom. Observation of the locomotion was classified into either 1-present or 2-absent. Observation of the sedation degree was classified as either 1-light (gets up when stimulated), 2-moderate (answers to incentives but stays in decubitus), or 3-intense (does not answer to incentives). The time of coagulation was determined using the capillary method (Jain 1993) after collection of blood from the tail. Coagulation was classified as either 1-coagulated or 2-not coagulated.

Hematology: Animals were subjected to euthanasia by the end of each period using 150mg/kg of intraperitoneal pentobarbital. Euthanasia was conducted soon after the intracardiac collection of blood. Total counting of erythrocyte and hemoglobin dosage for the cyanometahemoglobin method was performed by electronically counting the cells (CELM-CC530). Globular volume for the microhematocrit method was performed using a personal computer EULAB, model 024. The total plasma protein for refractometry (AST-Japan), fibrinogen for the precipitation technique for heat analysis (Kaneko *et al.* 1997), counting of leukocytes and differential counting of leukocytes was accomplished in 100 cells in a smear stained with fast Panotico (LABOR-CLIN) (Jain 1993).

Serum chemistry: Urea (Kit Labest-Lagoa Santa-MG), creatinine (Kit Bioclin-Belo Horizonte-MG), creatine kinase (CK)(Kit Doles-Goiânia-GO) (Kaneko *et al.* 1997).

Statistical analysis: Parametric data were subjected to variance analysis. Non-parametric data were subjected to Kruskal-Wallis analysis (Curi 1997).

RESULTS

Clinical exam: After venom inoculation, all animals presented discomfort, uneasiness, ataxia, pain and retraction of the limb where inoculation took place. Animals from groups VS and VSM presented edema of both rear limbs, with the left one being more swollen. Animals from group VS presented statistically significant edema ($p<0.05$) at M1, M2 and M3 when compared to the control group, and in group VSM, edema was more noticeable at M1. Intense edema was noticed in animals from group VS when compared to VSM (Table 1).

A significant drop in temperature ($p<0.05$) was noticed after six hours of inoculation for animals of groups VS and VSM, when compared to the control, and the values returned closer to the control group after 24 hours. In spite of not being statistically significant, a larger decrease in heart frequency was observed at two periods (M2 and M3) for group VS compared with the decrease at one period (M3) for group VSM (Table 1).

Moderate sedation at period M2 was statistically significant ($p<0.05$) for groups VS and VSM compared to group C. The animals of group VSM totally recovered at period M3, faster than the animals of group VS for the same period of time (Table 1).

The locomotion diminution was significant ($p<0.05$) at M2 for groups VS and VSM when compared to group C, and by clinical observation, animals from group VSM recovered faster than those of group VS (Table 1).

Laboratory analysis: Blood incoagulability was only observed in two animals; one from group VS at M1, and another from group VSM at M3. Thus, this information was neither statistically or clinically significant.

The number of red blood cells, hemoglobin and globular volume diminished at all periods for all animals from groups VS and VSM compared to the control group. Nevertheless,

TABLE 1

Average and standard deviation of the values found in the clinical exam in the different groups at different periods

Variable	Groups	Periods		
		M1 = 30 minutes after inoculation	M2 = 6 hours after inoculation	M3 = 24 hours after inoculation
Edema (member inoculated with venom)	C	0.77 ± 0.10	0.88 ± 0.16	0.77 ± 0.08
	VS	1.32 ± 0.12 ^A	1.53 ± 0.20 ^A	1.15 ± 0.29 ^A
	VSM	1.33 ± 0.16 ^A	1.08 ± 0.13 ^A	1.28 ± 0.10 ^A
Temperature	C	37.1 ± 0.2	37.2 ± 0.3	36.6 ± 0.8
	VS	37.3 ± 0.7	34.6 ± 1.6 ^A	35.8 ± 0.9
	VSM	36.1 ± 1.7	35.2 ± 1.1 ^A	35.8 ± 0.9
Respiratory frequency	C	104.3 ± 17.4	101.7 ± 17.6	116.3 ± 24.6
	VS	112.7 ± 25	90.3 ± 16.4	121.0 ± 8.9
	VSM	96 ± 17.2	119.7 ± 25.1	119.2 ± 18.1
Heart frequency	C	218 ± 11.2	218 ± 13.1	210 ± 14.3
	VS	212 ± 18.7	202 ± 43.1	126 ± 77.2
	VSM	213 ± 17.8	210 ± 15.1	179 ± 30.8
Sedation	C	1.0 (1.0;1.0)	1.0 (1.0;1.0)	1.0 (1.0;1.0)
	VS	1.0 (1.0;1.0)	2.0 (1.0;2.0) ^{AB}	1.5 (1.0;2.0) ^A
	VSM	1.0 (1.0;1.0)	2.0 (2.0;2.0) ^{AB}	1.0 (1.0;1.0)
Locomotion	C	1.0 (1.0;1.0)	1.0 (1.0;1.0)	1.0 (1.0;1.0)
	VS	1.0 (1.0;1.0)	2.0 (1.0;2.0) ^{AB}	1.0 (1.0;2.0)
	VSM	1.0 (1.0;1.0)	2.0 (1.0;2.0) ^{AB}	1.0 (1.0;1.0)

A= p<0.05 compare groups. B= p<0.05 compare periods.

C-control; VS-venom and antivenom; VSM-venom, antivenom and aqueous extract of *M. glomerata*.

Units are as follows: Rear left limb edema: cm. Temperature: degrees celsius. Respiratory frequency: movements per minute (mpm). Heart frequency: beats per minute (bpm).

Normal values: Temperature=35.9-37.5°C; Respiratory frequency=70-120mpm; Heart frequency=210-450bpm.

this was much more evident in group VS than in group VSM (Table 2).

An increase in the number of leukocytes and neutrophils (p<0.05) was observed in groups VS and VSM at M1 and M2 when compared to group C. Within the same group, when comparing among the observed time periods, neutrophil increase was larger (p<0.05) at period M2 than M1; while, lymphocytes were reduced for groups VS and VSM at M2, and for group VSM at M3 (Table 2).

Total plasma protein, fibrinogen, urea and creatinine values were within regular values for the species. Creatinine kinase levels were significantly higher (p<0.05) for groups VS and VSM at M1 and M2 compared with the control group. Values returned to normal for both groups at M3, but resulted higher for the animals of VS group (Table 3).

DISCUSSION

Edema happens due to direct poison action in muscles, and it has systemic effects on crotoxin, phospholipase A2, and several other substances that are released in the acute inflammatory phase such as interleukin, bradikinin and histamine (Kini & Evans 1989, Barraviera 1994). The results that were observed in this study agree with other authors that mention discreet to moderate edema at the site of inoculation of the venom (Hudelson & Hudelson 1995, Collicchio *et al.* 2002, Nogueira *et al.* 2007).

Animals of group VSM presented less intense edema in the member where the venom was inoculated, due to the action of the *M. glomerata*, agreeing with the results of Maiorano *et al.* (2005), that the plant can inhibit the activity of phospholipase A2 in mice. In

TABLE 2

Average and standard deviation of the values found in the hematology in the different groups at different periods

Variable	Groups	Periods		
		M1 = 30 minutes after inoculation	M2 = 6 hours after inoculation	M3 = 24 hours after inoculation
Red blood cells	C	8.11±469.65	6.93±389.05	6.89±261.30
	VS	8.64±623.56	6.27±778.08	7.28±838.97
	VSM	10.37±1,007.08	8.17±635.94	8.74±482.86
Hemoglobin	C	14.58±0.2	13.95±0.6	14.25±0.42
	VS	17.57±0.4	16.33±1.38	14.95±0.83
	VSM	19.56±1.0	16.35±0.1	15.77±0.8
Globular Volume	C	43.50±1.04	42.17±1.32	42.50±0.54
	VS	53.67±1.96	49.00±3.68	45.00±2.68
	VSM	55.23±3.87	49.17±1.32	47.83±2.4
Leukocytes	C	5.33±1,087.5	5.18±670.57	5.36± 939.5
	VS	10.08±3,185.9 ^{AB}	8.33±2,739.8 ^{AB}	3.58±1,049.6
	VSM	9.88±2,472.6 ^{AB}	14.01±5,093.7 ^{AB}	5.30±2,383.3
Neutrophils	C	2.29±548.48	1.78±380.08	2.01±205.83
	VS	4.87±1,180.7 ^{AB}	6.74±2,195.4 ^{AB}	1.66±517.87
	VSM	3.86±1,060.2 ^A	11.07±4,591.3 ^{AB}	4.03±1,825.7
Lymphocytes	C	2.56±490.95	3.07±574.40	2.73±871.64
	VS	4.27±2,179.6	1.16±548.71	1.57±480.66
	VSM	5.16±1,726.5	2.37±696.82	2.83±710.10
Total plasma protein	C	6.40±0.12	6.10±0.27	6.90±0.20
	VS	6.30±0.41	5.83±0.34	7.53±0.35
	VSM	6.267±0.37	6.33±0.30	6.53±0.56
Fibrinogen	C	300.00±109.54	233.33±81.650	266.67±103.28
	VS	200.00±0	300.00 ±109.54	366.67±150.55
	VSM	233.33±81.65	266.67±103.28	366.67±81.33

A= p<0.05 compare groups. B= p<0.05 compare periods.

C-control; VS-venom and antivenom; VSM-venom, antivenom and aqueous extract of *M.glomerata*.

Units are as follows. Red blood cells: $\times 10^6/\text{mm}^3$. Hemoglobin and total plasma protein: g/dL. Globular volume: %. Leukocytes, neutrophils and lymphocytes: $\times 10^3/\text{mm}^3$. Fibrinogen: mg/dL.

the present study, only the leaves were used for preparation of the plant extract, and in the study accomplished by Maiorano *et al.* (2005), the leaves, stalks and roots were used, which inhibited 100% the phospholipase A2 activity.

Temperature decrease occurred due to the state of shock and release of vasoactive agents at anaphylaxis, which is consistent with other reports, including hypothermia between six and 24 hours after venom inoculation (Lago *et al.* 2000, Raiser 2002, Nogueira & Sakate 2006).

Dyspnea and lower respiratory and cardiac frequencies were seen, especially in animals

from group VS, which are related to the cro-talic venom's myotoxic and neurotoxic action. This goes along with findings from Amaral *et al.* (1991), Nogueira & Sakate (2006) and Nogueira *et al.* (2007).

Sedation, diminution of locomotion and ataxia occurs due to the action of the cro-toxin and crotamine on the central and out-lying nervous system (Araújo & Belluomini 1962, Azevedo-Marques *et al.* 1985, Barraviera 1994, Vieira *et al.* 2006). The results from this study are in accordance with the findings from other authors that report the same clinical signs

TABLE 3

Average and standard deviation of the values found in the serum biochemistry in the different groups at different periods

Variable	Groups	Periods		
		M1 = 30 minutes after inoculation	M2 = 6 hours after inoculation	M3 = 24 hours after inoculation
Urea	C	39.90±15.6	44.90±11.7	47.68±1.85
	VS	47.68±1.85	60.70±9.29	41.17±5.27
	VSM	67.75±5.7	62.60±9.8	83.02±47.59
Creatinine	C	0.59±0.52	0.57±0.57	0.46±0.46
	VS	0.94±0.07	0.59±0.02	0.62±0.14
	VSM	0.75±0.1	0.55±0.07	0.66±0.06
Creatinine kinase	C	38.92±24	60.50±36.65	68.05±33.42
	VS	477.42±195.59 ^{AB}	385.08±206.17 ^{AB}	68.05±33.42
	VSM	378.9±131.01 ^{AB}	269.35±104.22 ^{AB}	76.72±78.62

A= p<0.05 compare groups. B= p<0.05 compare periods.

C-control; VS–venom and antivenom; VSM–venom, antivenom and aqueous extract of *M. glomerata*.

Units are as follows. Urea and creatinine: mg/dL. Creatinine kinase: UI/L.

in bovine two to six hours after venom inoculation (Belluomini *et al.* 1982, Lago *et al.* 2000), and in dogs two, four and six hours after venom inoculation (Collicchio *et al.* 2002, Nogueira & Sakate 2006, Nogueira *et al.* 2007). Our study did not show a significant difference between groups VS and VSM, thus differing from the results obtained by Maiorano *et al.* (2005), in which the venom's neurotoxic action was completely inhibited by extracts from fresh or dried roots and stems of *M. glomerata*. This difference was probably due to the use of a different part of the plant to treat the animals.

Blood incoagulability happened in a few animals but was neither statistically or clinically significant, probably due to the low sensitivity that this species has to crotalic venom. However, Maiorano *et al.* (2005) reported results with *M. glomerata* that showed that its extracts act as powerful inhibitors of clotting activity, probably due to interactions with thrombin-like enzymes.

The diminution in red blood cells, globular volume and hemoglobin after crotalic experimental envenomation is consistent with the hematological findings from Nogueira *et al.* (2007) and Nogueira & Sakate (2006), who observed the same results in dogs.

Leukocytosis with neutrophilia that was observed in groups VS and VSM at M1 and

M2 in this study, together with lymphopenia at M2 and M3 in both groups, are consistent with other authors, and this may be due to inflammatory processes and stress caused by general pain (Santoro *et al.* 2008, Ferreira Junior & Barraviera 2004, Nogueira *et al.* 2007).

The average total plasma protein, fibrinogen, urea and creatinine values were within reference values for the species and no differences among treatments were observed.

Crotalic venom has a myotoxic action (Nogueira *et al.* 2007, Azevedo-Marques *et al.* 1985, Bucarechi *et al.* 2002, Ferreira Junior & Barraviera 2004), and creatine kinase is important in identifying muscle lesions (Kaneko *et al.* 1997). The myotoxic action promotes widespread myalgia and myoglobinuria (Azevedo-Marques *et al.* 1985, Magalhães *et al.* 1986, Motta *et al.* 2007).

This study showed an increase in CK values for groups VS and VSM at M1 and M2, consistent with the authors that mention that crotalic venom has effects on muscle fibers (Azevedo-Marques *et al.* 1985, Lago *et al.* 2000).

There was no statistical difference (p>0.05) among treatments, but the values of CK were higher in animals of group VS. The aqueous extract of *M. glomerata* might have lowered CK values for animals of group VSM by

inhibiting the action of phospholipase A2, as described by Maiorano *et al.* (2005). A deeper investigation with histological analysis of the fabric of the member inoculated is necessary.

RESUMEN

El envenamiento crotálico representa el número más alto de muertes cuando es comparado con envenenamientos por mordeduras de otras serpientes de interés médico. El veneno crotálico tiene importantes características de acción neurotóxica, miotoxicidad, nefrotoxicidad, coagulación y acción hemolítica. Este trabajo evaluó los aspectos clínicos y de laboratorio del envenenamiento experimental con el veneno de la serpiente *Crotalus durissus terrificus* en las ratas Wistar tratadas con suero antiofídico y extracto acuoso de *M. glomerata*. Los animales fueron separados en tres diferentes grupos: grupo control (C); grupo veneno+suero (VS), grupo veneno+suero+extracto acuoso de *M. glomerata* (VSM). El veneno crotálico causó alteraciones clínicas y diferencias en los análisis sanguíneos practicados a los ratones Wistar evaluados. Las alteraciones clínicas más importantes fueron una disminución de la temperatura, edema en el miembro inoculado de veneno, la sedación y una disminución de la locomoción en los grupos VS y VSM comparado con el grupo C. Una rápida recuperación de la sedación estadísticamente significativa fue observada en los animales del grupo VSM al compararse con los del grupo VS. Los análisis sanguíneos mostraron un aumento en el número de leucocitos, neutrofilos y creatina quinasa en los grupos VS y VSM comparados con el grupo C. Los ratones Wistar mostraron una alta resistencia al veneno del crotálico. Estudios adicionales con variación en las dosis, tiempo de tratamiento, y métodos de administración, así como la realización de estudios histopatológicos e inmunológicos son importantes para comprender la acción de *M. glomerata* en accidentes crotálicos.

Palabras clave: *Crotalus durissus terrificus*, *Mikania glomerata*, suero antiofídico, rata, veneno

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